

Epstein-Barr Virus (VCA)

IgM – ELISA

Enzyme immunoassay for the qualitative determination of IgM-class antibodies against Epstein-Barr Virus in human serum or plasma

For laboratory research only.

Product Number: NB-06-0519 (96 Determinations)



1. INTRODUCTION

Epstein-Barr Virus (EBV) is a member of the herpesvirus family (Gamma subgroup, DNA virus of 120-200 nm) and one of the most common human viruses. The virus occurs worldwide, and most people become infected with EBV sometime during their lives. Transmission of the virus is almost impossible to prevent since many healthy people can carry and spread the virus intermittently for life. Infants become susceptible to EBV as soon as maternal antibody protection disappears. Infection of children usually causes no symptoms. Infection during adolescence or young adulthood causes infectious mononucleosis 35% to 50% of the time.

Infectious mononucleosis is almost never fatal. There are no known associations between active EBV infection and problems during pregnancy, such as miscarriages or birth defects. Although the symptoms of infectious mononucleosis usually resolve in 1 or 2 months, EBV remains dormant or latent in a few cells in the throat and blood for the rest of the person's life. Periodically, the virus can reactivate and is commonly found in the saliva of infected persons. This reactivation usually occurs without symptoms of illness. EBV also establishes a lifelong dormant infection in some cells of the body's immune system. A late event in a very few carriers of this virus is the emergence of Burkitt's lymphoma and nasopharyngeal carcinoma, but EBV is probably not the sole cause of these malignancies.

Species	Disease	Symptoms	Mechanism of Infection
Epstein-Barr	infectious mononucleosis	fever, sore throat, swollen lymph	Person to Person Transmission
Virus		glands	
			EBV requires intimate contact with the saliva of
			an infected person, but the virus is also found in
			the saliva of healthy people

The presence of virus resp. infection may be identified by

- PCR
- Serology: "mono spot" test, Detection of antibodies by ELISA

The optimal combination of serologic testing consists of the titration of four markers: IgM and IgG to the viral capsid antigen (VCA), IgM to the early antigen, and antibody to EBV nuclear antigen (EBNA). IgM to VCA appears early in infection and disappears within 4 to 12 weeks. IgG to VCA appears in the acute phase, peaks at 2 to 4 weeks after onset, declines slightly, and then persists for life. If antibodies to the viral capsid antigen are not detected, the patient is susceptible to EBV infection.

2. INTENDED USE

The NeoBiotech Epste in-Barr Virus (EBV) IgM-ELISA is intended for the qualitative determination of IgM class antibodies against Epstein-Barr virus **viral capsid antigen (VCA)** in human serum or plasma (citrate).

3. PRINCIPLE OF THE ASSAY

The qualitative immunoenzymatic determination of IgM-class antibodies against Epstein-Barr Virus is based on the ELISA (Enzymelinked Immunosorbent Assay) technique.

Microtiter strip wells are precoated with Epstein-Barr Virus antigens to bind corresponding antibodies of the specimen. After washing the wells to remove all unbound sample material horseradish peroxidase (HRP) labelled anti-human IgM conjugate is added. This conjugate binds to the captured Epstein-Barr Virus-specific antibodies. The immune complex formed by the bound conjugate is visualized by adding Tetramethylbenzidine- (TMB) substrate which gives a blue reaction product. The intensity of this product is proportional to the amount of Epstein-Barr Virus-specific IgM antibodies in the specimen. Sulphuric acid is added to stop the reaction. This produces a yellow endpoint colour. Absorbance at 450 nm is read using an ELISA microwell plate reader.

4. MATERIALS

4.1. Reagents supplied

- Epstein-Barr Virus Coated Wells (IgM): 12 breakapart 8-well snap-off strips coated with Epstein-Barr Virus antigen; in resealable aluminium foil.
- **IgM Sample Diluent** ***: 1 bottle containing 100 ml of buffer for sample dilution; containing antihuman-IgG; pH 7.2 ± 0.2; coloured green; ready to use; white cap.
- Stop Solution: 1 bottle containing 15 ml sulphuric acid, 0.2 mol/l; ready to use; red cap.
- Washing Solution (20x conc.)*: 1 bottle containing 50 ml of a 20-fold concentrated buffer (pH 7.2 ± 0.2) for washing the wells; white cap.
- Epstein-Barr Virus anti-IgM Conjugate**: 1 bottle containing 20 ml of peroxidase labelled rabbit antibody to human IgM; coloured red, ready to use; black cap.
- TMB Substrate Solution: 1 bottle containing 15 ml 3,3',5,5'-tetramethylbenzidine (TMB); ready to use; yellow cap.
- Epstein-Barr Virus IgM Positive Control***: 1 bottle containing 2 ml; coloured yellow; ready to use; red cap.
- Epstein-Barr-Virus IgM Cut-off Control***: 1 bottle containing 3 ml; coloured yellow; ready to use; green cap.
- Epstein-Barr Virus IgM Negative Control***: 1 bottle containing 2 ml; coloured yellow; ready to use; blue cap.
- contains 0.1 % Bronidox L after dilution
- ** contains 0.2 % Bronidox L
- *** contains 0.1 % Kathon



4.2. Materials supplied

- 1 Strip holder
- 1 Cover foil
- 1 Test protocol
- 1 distribution and identification plan

4.3. Materials and Equipment needed

- ELISA microwell plate reader, equipped for the measurement of absorbance at 450/620nm
- Incubator 37°C
- Manual or automatic equipment for rinsing wells
- Pipettes to deliver volumes between 10 and 1000 ul
- Vortex tube mixer
- Deionised or (freshly) distilled water
- Disposable tubes
- Timer

5. STABILITY AND STORAGE

The reagents are stable up to the expiry date stated on the label when stored at 2...8 °C.

6. REAGENT PREPARATION

It is very important to bring all reagents, samples and controls to room temperature (20...25°C) before starting the test run!

6.1. Coated snap-off Strips

The ready to use breakapart snap-off strips are coated with Epstein-Barr Virus antigen. Store at 2...8°C. *Immediately after removal of strips, the remaining strips should be resealed in the aluminium foil along with the desiccant supplied and stored at 2...8°C; stability until expiry date.*

6.2. Epstein-Barr Virus anti-IgM Conjugate

The bottle contains 20 ml of a solution with anti-human-IgM horseradish peroxidase, buffer, stabilizers, preservatives and an inert red dye. The solution is ready to use. Store at 2...8°C. After first opening stability until expiry date when stored at 2...8°C.

6.3. Controls

The bottles labelled with Positive, Cut-off and Negative Control contain a ready to use control solution. It contains 0.1% Kathon and has to be stored at 2...8°C. After first opening stability until expiry date when stored at 2...8°C.

6.4. IgM Sample Diluent

The bottle contains 100 ml phosphate buffer, anti human-IgG, stabilizers, preservatives and an inert green dye. It is used for the dilution of the patient specimen. The solution contains antihuman IgG class antibodies to eliminate competitive inhibition from specific IgG class antibody to remove rheumatoid factor. This ready to use solution has to be stored at 2...8°C. After first opening stability until expiry date when stored at 2...8°C.

6.5. Washing Solution (20xconc.)

The bottle contains 50 ml of a concentrated buffer, detergents and preservatives. Dilute washing solution 1+19; e.g. 10 ml washing solution + 190 ml fresh and germ free redistilled water. The diluted buffer will keep for 5 days if stored at room temperature. *Crystals in the solution disappear by warming up to 37 °C in a water bath. After first opening the concentrate is stable until the expiry date.*

6.6. TMB Substrate Solution

The bottle contains 15 ml of a tetramethylbenzidine/hydrogen peroxide system. The reagent is ready to use and has to be stored at 2...8°C, away from the light. The solution should be colourless or could have a slight blue tinge. If the substrate turns into blue, it may have become contaminated and should be thrown away. After first opening stability until expiry date when stored at 2...8°C.

6.7. Stop Solution

The bottle contains 15 ml 0.2 M sulphuric acid solution (R 36/38, S 26). This ready to use solution has to be stored at 2...8°C. After first opening stability until expiry date.

7. SPECIMEN COLLECTION AND PREPARATION

Use human serum or plasma (citrate) samples with this assay. If the assay is performed within 5 days after sample collection, the specimen should be kept at 2...8°C; otherwise they should be aliquoted and stored deep-frozen (-20 to -70°C). If samples are stored frozen, mix thawed samples well before testing. Avoid repeated freezing and thawing. Heat inactivation of samples is not recommended.



7.1. Sample Dilution

Before assaying, all samples should be diluted 1+100 with IgM Sample Diluent. Dispense 10μ l sample and 1 ml IgM Sample Diluent into tubes to obtain a 1+100 dilution and thoroughly mix with a Vortex.

8. ASSAY PROCEDURE

8.1. Test Preparation

Please read the test protocol carefully **before** performing the assay. Result reliability depends on strict adherence to the test protocol as described. The following test procedure is only validated for manual procedure. If performing the test on ELISA automatic systems we recommend to increase the washing steps from three to five and the volume of washing solution from 300µl to 350µl to avoid washing effects. Prior to commencing the assay, the distribution and identification plan for all specimens and controls should be carefully established on the result sheet supplied in the kit. Select the required number of microtiter strips or wells and insert them into the holder.

Please allocate at least:

1 well (e.g. A1) for the substrate blank, 1 well (e.g. B1) for the negative control, 2 wells (e.g. C1+D1) for the cut-off control and 1 well (e.g. E1) for the positive control.

It is recommended to determine controls and patient samples in duplicate.

Perform all assay steps in the order given and without any appreciable delays between the steps.

A clean, disposable tip should be used for dispensing each control and sample.

Adjust the incubator to $37^{\circ} \pm 1^{\circ}$ C.

- 1. Dispense 100µl controls and diluted samples into their respective wells. Leave well A1 for substrate blank.
- 2. Cover wells with the foil supplied in the kit.
- 3. Incubate for 1 hour \pm 5 min at $37\pm1^{\circ}$ C.
- 4. When incubation has been completed, remove the foil, aspirate the content of the wells and wash each well three times with 300μl of Washing Solution. Avoid overflows from the reaction wells. The soak time between each wash cycle should be >5sec. At the end carefully remove remaining fluid by tapping strips on tissue paper prior to the next step!
 - Note: Washing is critical! Insufficient washing results in poor precision and falsely elevated absorbance values.
- 5. Dispense 100µl Epstein-Barr Virus anti-IgM Conjugate into all wells except for the blank well (e.g. A1). Cover with foil.
- 6. **Incubate for 30 min at room temperature.** *Do not expose to direct sunlight.*
- 7. Repeat step 4.

Note:

- 8. Dispense 100µl TMB Substrate Solution into all wells
- 9. Incubate for exactly 15 min at room temperature in the dark.
- Dispense 100μl Stop Solution into all wells in the same order and at the same rate as for the TMB Substrate Solution.
 Any blue colour developed during the incubation turns into yellow.

Highly positive patient samples can cause dark precipitates of the chromogen! These precipitates have an influence when reading the optical density. Predilution of the sample with physiological sodium chloride solution, for example 1+1, is recommended. Then dilute the sample1+100 with dilution buffer and multiply the results in NTU by 2.

11. Measure the absorbance of the specimen at 450/620nm within 30 min after addition of the Stop Solution.

8.2. Measurement

Adjust the ELISA Microwell Plate Reader to zero using the substrate blank in well A1.

If - due to technical reasons - the ELISA reader cannot be adjusted to zero using the substrate blank in well A1, subtract the absorbance value of well A1 from all other absorbance values measured in order to obtain reliable results!

Measure the absorbance of all wells at 450 nm and record the absorbance values for each control and patient sample in the distribution and identification plan.

Dual wavelength reading using 620 nm as reference wavelength is recommended.

Where applicable calculate the mean absorbance values of all duplicates.

9. RESULTS

9.1. Run Validation Criteria

In order for an assay to be considered valid, the following criteria must be met:

■ **Substrate blank** in A1: Absorbance value < **0.100.**

• Negative control in B1: Absorbance value < 0.200 and < cut-off



Cut-off control in C1 and D1: Absorbance value 0.150 – 1.30.
 Positive control in E1: Absorbance value > cut-off.

If these criteria are not met, the test is not valid and must be repeated.

9.2. Calculation of Results

The cut-off is the mean absorbance value of the Cut-off control determinations.

Example: Absorbance value Cut-off control 0.39 + absorbance value Cut-off control 0.37 = 0.76 / 2 = 0.38

Cut-off = 0.38

9.3. Interpretation of Results

Samples are considered **POSITIVE** if the absorbance value is higher than 10% over the cut-off.

Samples with an absorbance value of 10% above or below the cut-off should not be considered as clearly positive or negative → grey zone

It is recommended to repeat the test again 2 - 4 weeks later with a fresh sample. If results in the second test are again in the grey zone the sample has to be considered **NEGATIVE**.

Samples are considered **NEGATIVE** if the absorbance value is lower than 10% below the cut-off.

9.3.1. Results in NeoBiotech Units

Patient (mea	ın) absorbance value	e x 10	=	[NeoBiotech-Units = NTU]
	Cut-off			
Example:	$1.786 \times 10 = 47$	NTU		
	0.38			
Cut-off:	10	NTU		
Grey zone:	9-11	NTU		
Negative:	<9	NTU		
Positive:	>11	NTU		

10. SPECIFIC PERFORMANCE CHARACTERISTICS

10.1. Precision

Interassay	n	Mean	Cv (%)
Pos. Serum	24	1.37	6.9
Intraassay	n	Mean	Cv (%)
Pos. Serum	25	1.32	5.1

10.2. Diagnostic Specificity

The diagnostic specificity is defined as the probability of the assay of scoring negative in the absence of the specific analyte. It is 96.3 %.

10.3. Diagnostic Sensitivity

The diagnostic sensitivity is defined as the probability of the assay of scoring positive in the presence of the specific analyte. It is 93.9 %.

10.4. Interferences

Interferences with hemolytic, lipemic or icteric sera are not observed up to a concentration of 10 mg/ml hemoglobin, 5 mg/ml triglycerides and 0.2 mg/ml bilirubin.

Panel Prof. Krech/Frauenfeld/Swiss



10.5. Cross-Reactivity

Acute Infection	EBV IgM ELISA			
Adenovirus	neg.			
CMV	neg.			
Echinococcus	neg.			
HBV	neg.			
Influenza A	neg.			
Influenza B	neg.			
Leptospira	neg.			
Picorna	neg.			
Q-Fever	neg.			
Rubella neg.				
Toxoplasma	neg.			
VZV	neg.			
Lues	neg.			
RSV	neg.			

Note: The results refer to the groups of samples investigated; these are not guaranteed specifications.

11. LIMITATIONS OF THE PROCEDURE

Bacterial contamination or repeated freeze-thaw cycles of the specimen may affect the absorbance values. Diagnosis of an infectious disease should not be established on the basis of a single test result. A precise diagnosis should take into consideration clinical history, symptomatology as well as serological data.

In immunocompromised patients and newborns serological data only have restricted value.

12. PRECAUTIONS AND WARNINGS

- In compliance with article 1 paragraph 2b European directive 98/79/EC the use of the in vitro diagnostic medical devices is intended by the manufacturer to secure suitability, performances and safety of the product. Therefore the test procedure, the information, the precautions and warnings in the instructions for use have to be strictly followed. The use of the testkits with analyzers and similar equipment has to be validated. Any change in design, composition and test procedure as well as for any use in combination with other products not approved by the manufacturer is not authorized; the user himself is responsible for such changes. The manufacturer is not liable for false results and incidents for these reasons. The manufacturer is not liable for any results by visual analysis of the patient samples.
- Only for in-vitro diagnostic use.
- All components of human origin used for the production of these reagents have been tested for <u>anti-HIV antibodies</u>, <u>anti-HCV antibodies and HBsAg and have been found to be non-reactive</u>. Nevertheless, all materials should still be regarded and handled as potentially infectious.
- Do not interchange reagents or strips of different production lots.
- No reagents of other manufacturers should be used along with reagents of this test kit.
- Do not use reagents after expiry date stated on the label.
- Use only clean pipette tips, dispensers, and lab ware.
- Do not interchange screw caps of reagent vials to avoid cross-contamination.
- Close reagent vials tightly immediately after use to avoid evaporation and microbial contamination.
- After first opening and subsequent storage check conjugate and control vials for microbial contamination prior to further use.
- To avoid cross-contamination and falsely elevated results pipette patient samples and dispense conjugate without splashing accurately to the bottom of wells.
- The NeoBiotech ELISA is only designed for qualified personnel who are familiar with good laboratory practice.

WARNING:	In the used concentration Bronidox L has hardly any toxicological risk upon contact with skin and mucous membranes!
WARNING:	Sulphuric acid irritates eyes and skin. Keep out of the reach of children. Upon contact with the eyes, rinse thoroughly with water and consult a doctor!

12.1. Disposal Considerations

Residues of chemicals and preparations are generally considered as hazardous waste. The disposal of this kind of waste is regulated through national and regional laws and regulations. Contact your local authorities or waste management companies which will give advice on how to dispose hazardous waste.

13. ORDERING INFORMATION

Prod. No.: NB-06-0519 Epstein-Barr Virus (VCA) IgM-ELISA (96 Determinations)



BIBLIOGRAPHY

Bergman M., Gleckman R.A., "Heterophil-negative infectious mononucleosis-like syndrome". Postgrad.Med., 81 (1): 313-326 (1987) Buchwald D., KOmaroff A.L., "Review of laboratory findings for patient with chronic fatique syndrome". Rev. Inf. Dis., 13 (Suppl. 1): S12-S18 (1991)

De Ory F., Antonaya J., Fernandez M.V., Echevarria J.M., "Application of low-avidity immunglobulin G studies to diagnosis of EBV infectious mononucleosis". J. Clin. Microbiol., 31 (6): 1669-1671, (1993)

De-The, G., "Epidemiology of EBV and Associated Diseases in Man", In: The Herpesviruses, Roizman, B. (ed)., Volume 1, New York: Plenum-Press, 25-103, (1982)

Dölken G., Weitzmann U., Boldt C. et al.. "Enzyme linked immunosorbent assay for IgG antibodies of EBV virus associated early antigens and viral capside antigen".J. Immunol. Methz., 67: 225-233, (1984)

Färber I., Wutzler P., Wohlrabe P. et al., "Serological diagnosis of infectious mononucleosis using three anti-Epstein-Barr virus recombinant ELISAs", J. Virol. Meth.; 42: 301-308, (1993)

Gorgievski-Hrisoho M., Hinderer W., Nebel-Schickel H. et al., "Serodiagnosis of infectious mononucleosis by using recombinant Epstein-Barr virus antigens and enzyme-linked Immunosorbent assay technology", J. Clin. Microbiol., 28 (10): 2305-2311, (1990)

Halprin J., Scott A.L., Jacoboson L. et al., "Enzyme-linked immunosorbent assay of antibodies to Epstein-Barr virus nuclear and early antigens in patients with infectious mononucleosis and nasopharyngeal carcinoma", Ann. Int. Med., 104: 331-337, (1986).

Heath, C.W., A.L. Brodsky, and A.L. Ptolosky, "Infectious Mononuclelosis in a general population", Am. J. Epiemiology, 95 (1): 46-52, (1972)

Henle W., Henle GE., Horwitz CA., "Epstein-Barr virus specific giagnostic tests in infectious mononucleosis", Human Pathol., 5 (5): 551-565, (1974)

Lamy ME., Favart AM., Cornu A. et al., "Study of Epstein-Barr virus (EBV) antibodies: IgG and IgM anti-VCA, IgG anti EA and Ig anti-EBNA obtained with an original microtiter technique. Serological criterions of primary and recurrents EBV infections and follow-up of infectious mononucleosis. Seroepidemiology of EBV in Belgium based on 5178 sera from patients". Acta Clin. Belg., 37 (5): 281-298, (1982)

Lennette E., "Epstein-Barr Virus", in Manual of Clinical Microbiology, 4th ed. Washington D.C., Am. Soc. Microbiol. P 728-732, (1985)

Luka J., chase PC., Pearson GR., "A sensitive enzyme-linked Immunosorbent assay (ELISA) against the major EBV-associated antigens. I-Correlation between ELISA and immunofluorescence titers using purified antigens", J. Immunol. Metho., 67: 145-156, (1984)

Pearson GP., "Infectious Mononucleosis: the humoral response", In: Infectious mononucleosis, D. Schlossberg ed., Springer-Verlag, New York, p. 89-99, (1989)

Pocheldy C., "Laboratory testing for infectious mononucleosis: cautions to observe in interpreting results". Prostgrad. Med., 81 (1): 335-342 (1987)

Purtilo BT., Hinrichs S., "Detection of Epstein-Barr Virus induced deseases by laboratory techniques", Incstar Monograph, (1993)



Symbols Key				
W	Manufactured by			
IVD	In Vitro Diagnostic Medical Device			
LOT	Lot Number			
\square	Expiration Date			
*	Storage Temperature			
C€	CE Mark			
[REF]	Catalogue Number			
i	Consult Instructions for Use			
MTP	Microplate			
CONJ	Conjugate			
CONTROL -	Control serum, negative			
CONTROL +	Control serum, positive			
CUT OFF	Cut off control serum			
DIL M	Sample diluent buffer IgM			
SOLN STOP	Stop solution			
SUB TMB	TMB Substrate solution			
WASHBUF 20x	Washing solution 20x concentrated			
$\sum_{\mathbf{n}}$	Contains sufficient for "n" tests			

Epstein-Barr Virus IgM-ELISA

Test Preparation

Prepare reagents and samples as described.

Establish the distribution and identification plan for all specimens and controls on the result sheet supplied in the kit.

Select the required number of microtiter strips or wells and insert them into the holder.

	Substrate blank	Negative	Positive	Cut-off	Sample (diluted 1+100)	
	(e.g. A1)	control	control	control	(diluted 1+100)	
Negative control	-	100µl	-	-	-	
Positive control	-	-	100µl	-	-	
Cut-off control	-	-	-	100µl	-	
Sample (diluted 1+100)	-	-	-	-	100μ1	
	Cover wells with foil supplied in the kit					
	Iı	ncubate for 1	h at 37°C			
	Wash each well th	ree times with	a 300µl of was	shing solution		
Conjugate	-	100µl	100µ1	100µl	100µl	
Cover wells with foil supplied in the kit						
	Incubate f	or 30 min at	room temper	ature		
Wash each well three times with 300µl of washing solution						
TMB Substrate	100μ1	100µl	100µ1	100µl	100μ1	
Incubate for exactly 15 min at room temperature in the dark						
Stop Solution	100μ1	100µl	100µl	100µl	100μ1	
Photometric measurement at 450 nm (reference wavelength: 620 nm)						

engl07082009-JH